

Quantification and Qualification of Oligonucleotides by High Resolution/Accurate Mass Orbitrap MS

Hongxia (Jessica) Wang¹, Kevin Cook¹, Patrick Bennett¹

¹Thermo Fisher Scientific, San Jose, CA

Overview

Purpose: Identification and quantitation of oligonucleotides by UHPLC-high resolution Orbitrap MS.

Methods: Oligonucleotides were first qualified in full scan MS mode at 70,000 resolution, then quantified with both selected ion monitoring (SIM) at 70,000 resolution and targeted MS/MS methods at 17,500 resolution with a 8-min gradient on Thermo Hypersil Gold column.

Results: Samples prepared in a human plasma based solution to reduce non-specific binding indicates good linearity with a calibration range of 1-5nM to 10 μ M. Performing SIM (70,000 resolution) shows 5 fold more sensitivity than targeted MS/MS method (17,500 resolution). With faster scan speed, there are at least 20 scans across a 9-sec UHPLC peak around LOQ at 70,000 resolution.

Introduction

Qualitative and quantitative analysis of oligonucleotides in biological matrices are important aspects of the drug development process. There is a trend towards the use of high resolution mass spectrometry (HRMS) as an alternative to overcome the limitations of nominal resolution provided by triple quadrupole MS.¹⁻² A high throughput generic UHPLC-HR-MS assay is demonstrated for oligonucleotide analysis using Q-Exactive Orbitrap MS. The qualitative information provided during quantitation of analytes within one injection is a crucial advantage to accelerate drug development process by reducing instrument analysis time and sample consumption.

Methods

Sample Preparation

Oligonucleotides (ODNs)(15mer- and 20mer-),bought from TriLink BioTechnologies, were dissolved in water. The concentration of stock solutions were quantified by Thermo NanoDrop UV spectrometer. A 20-mer Phosphodiester ODN (5'-ATT CAG TTC ACT TAT CGT AT-3') was diluted in 3 different carriers (1)Solvent A (2%HFIP+0.4%TEA in Water) (2) 0.1% Human Plasma in Solvent A (3) 0.1% Human Plasma + 0.1%TFA+ 10%MeOH. The synthetic ODN was quantified with 15-mer phosphorothioate oligonucleotide (5'-ATT CAG TTC ACT TAT-3') as internal standard. Samples were analyzed on a Thermo Q-Exactive Orbitrap equipped with XRSTM UHPLC pump and Open Accela autosampler.

Liquid Chromatography

Column: Hypersil Gold C18 column (2.1 x 50 mm, 1.9 μ m)

Injection Volume: 10 μ L

LC: XRSTM UHPLC pump

Solvent A: 2%HFIP+0.4%TEA in Water

Solvent B: 2%HFIP+0.4%TEA in Methanol

Flow Rate: 300 μ L/min

Gradient:	Time	A%	B%
	0.0	90	10
	5.0	60	40
	6.0	10	90
	8.0	10	90
	8.1	95	10

Mass Spectrometry

Spray Voltage (-) 3800 kV

Capillary Temperature (-) 320 °C

Sheath Gas (-) 45

Aux Gas (-) 15

Sweep Gas (-) 0

Heater Temperature (-) 400 °C

S-lens 50

Negative MS Scan 1 microscan

Full Scan R = 70,000; AGC = 1e6; IT = 250 ms; Lock Mass = off

SIM R = 70,000; AGC=2e5; IT= 350ms; Isolation= 4 amu

MS/MS R = 17,500; AGC = 2e5; IT= 500 ms; Isolation= 4 amu;

HCD = 20

- Max resolution: 140,000 at m/z 200
- Scan speed: up to 12 Hz (at 17.5K)
- Mass Accuracy
 - < 5ppm external
 - < 1ppm internal
- Mass range for full scans: 50 < m/z < 6000
- Intra-scan dynamic range: > 5000:1
- Sensitivity
 - Full MS: 500 fg Buspirone on column S/N 100:1
 - SIM: 50 fg Buspirone on column S/N 100:1
- Polarity Switching
 - One full cycle in < 1 sec (one full scan positive mode and one full scan negative mode at resolution setting of 17,500)

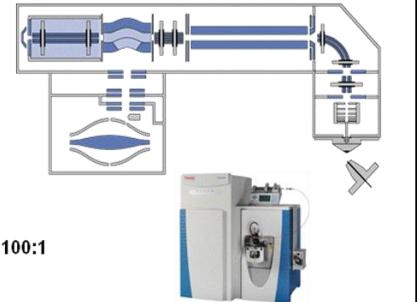


FIGURE 1. Q Exactive Benchtop Orbitrap Mass Spectrometer

Data Analysis

Oligonucleotides were initially analyzed by full scan mode at 70,000 resolution to obtain the charge state distribution envelope. The most abundant ions, $[M-9H]^{9-}$ for 20-mer and $[M-3H]^{3-}$ for 15-mer, were then employed for quantitation. A comparison of ODN quantitation using both selected ion monitoring (SIM) and targeted MS/MS was performed with a 8-min gradient to separate analyte with internal standard. The calibration curves of ODNs were generated by LCquan™ software. For SIM method, the four most abundant isotopes of deprotonated ODNs were summed for the quantitation. For targeted MS/MS, four relative abundant and unique fragmentation ions were chosen to do quantification.

Results

Oligonucleotides Charge State Distribution and SIM Quantitation

Analyte and internal standard ODNs were analyzed by LC-MS at full scan mode from m/z 400 to 2000 at 70,000 resolution. The charge state distributions of two ODNs are shown in Figure 2 and 4, respectively. For 20-mer ODN, the most abundant ion is $[M-9H]^{9-}$. Four isotopes, m/z 671.7792, 671.8902, 672.0001 and 672.1119 were selected for quantitation for SIM method (Figure 3). For 15-mer ODN, $[M-3H]^{3-}$ is the most abundant ion, three isotopes, m/z 1579.1551, 1579.4875 and 1579.8208 were used for quantitation (Figure 5).

FIGURE 2. Full Scan MS Spectrum of 20-mer Synthetic Oligonucleotide (5'-ATT CAG TTC ACT TAT CGT AT-3')

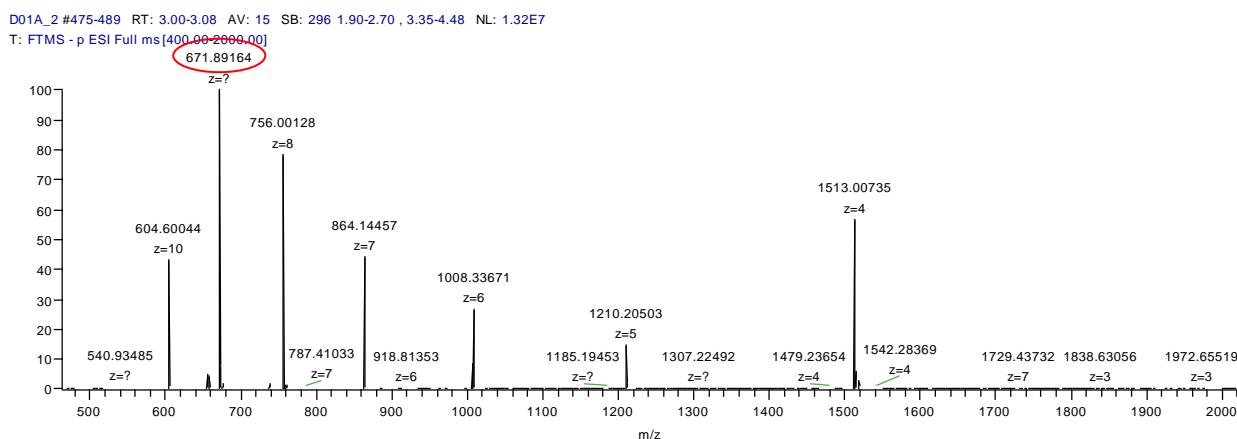


FIGURE 3. Zoom-in MS Spectrum of $[M-9H]^{9-}$ 20-mer Oligonucleotide 5'-ATT CAG TTC ACT TAT CGT AT-3')

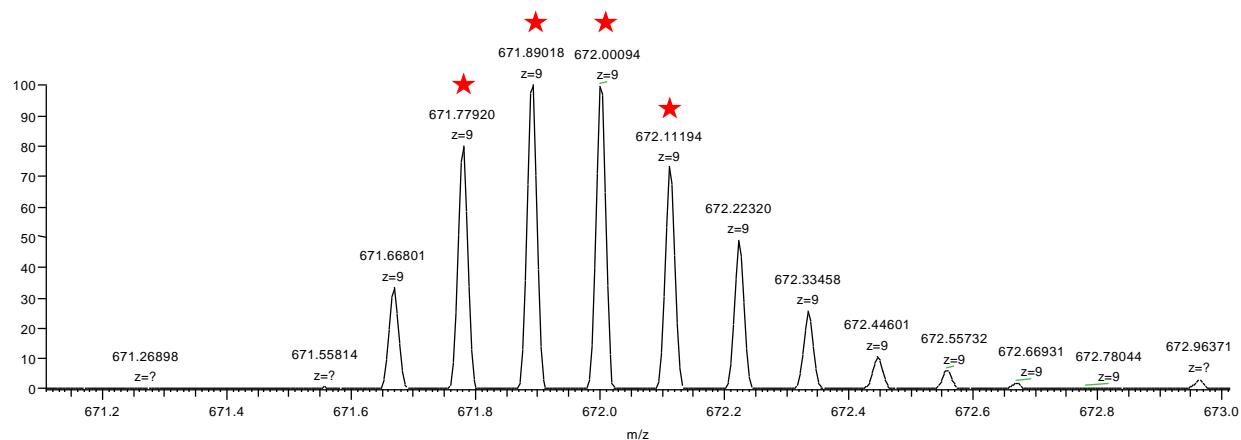


FIGURE 4. Full Scan MS Spectrum of 15-mer synthetic Oligonucleotide (5'-ATT CAG TTC ACT TAT-3')

15mer_1 #565-583 RT: 3.57-3.68 AV: 19 SB: 1502 0.23-3.15 , 4.69-10.72 NL: 4.47E6
T: FTMS - p ESI Full ms [400.00-2000.00]

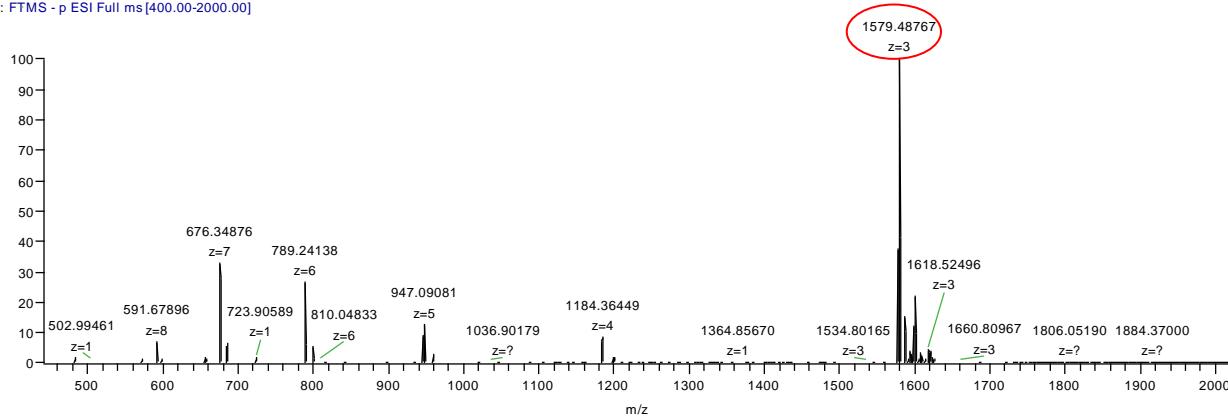
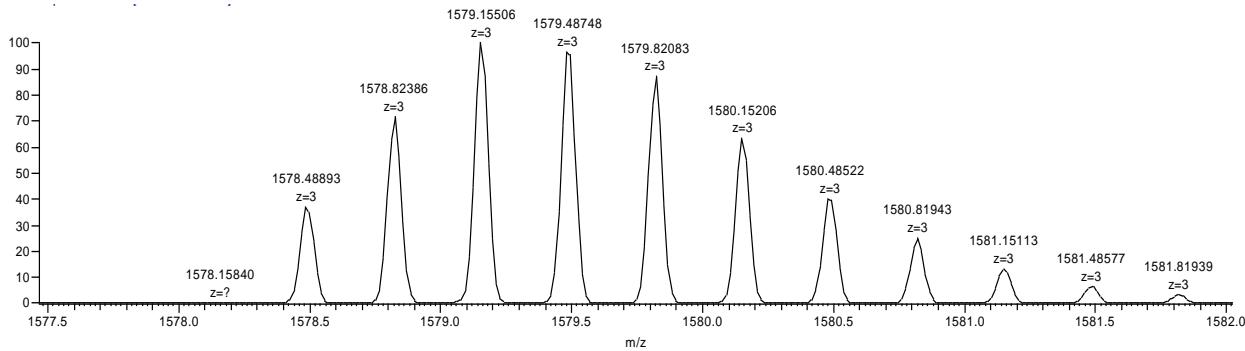


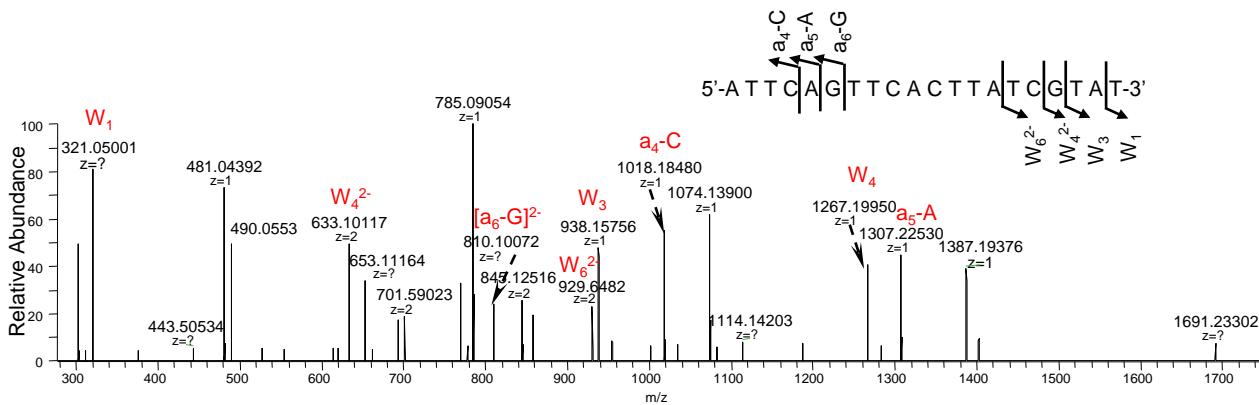
FIGURE 5. Zoom-in MS Spectrum for $[M-3H]^{3-}$ of 15-mer Oligonucleotide (5'-ATT CAG TTC ACT TAT-3')



Targeted MS/MS Quantitation

Four fragmentation ions, in Figure 6, m/z 938.1536, 1018.1786, 1267.2055 and 1387.1949 from $[M-4H]^{4-}$ (m/z 1513.0074) of 20-mer ODN were used for targeted MS/MS quantitation. While for 15-mer ODN, fragmentation ions m/z 968.0706, 833.0189, 1138.0447 and 1306.1107 from $[M-3H]^{3-}$ (m/z 1579.4876) precursor ion were chosen (spectrum not shown).

FIGURE 6. HCD MS/MS spectrum of $[M-4H]^{4-}$ (m/z 1512.0073) from 20-mer ODN (CE=20)



Calibration Curve

To avoid analyte absorption, three different carriers (see Method) and two sample vials (polypropylene and silanized glass) were evaluated. Silanized glass vials and human plasma in 10% MeOH were found to give the least absorption. ODN (20-mer) was then quantified by both SIM and MS/MS methods. SIM method (70,000 resolution) shows 5 fold more sensitivity than the targeted MS/MS method (17,500 resolution) (Figure 7).

FIGURE 7. SIM and MS/MS Chromatograms near LOD

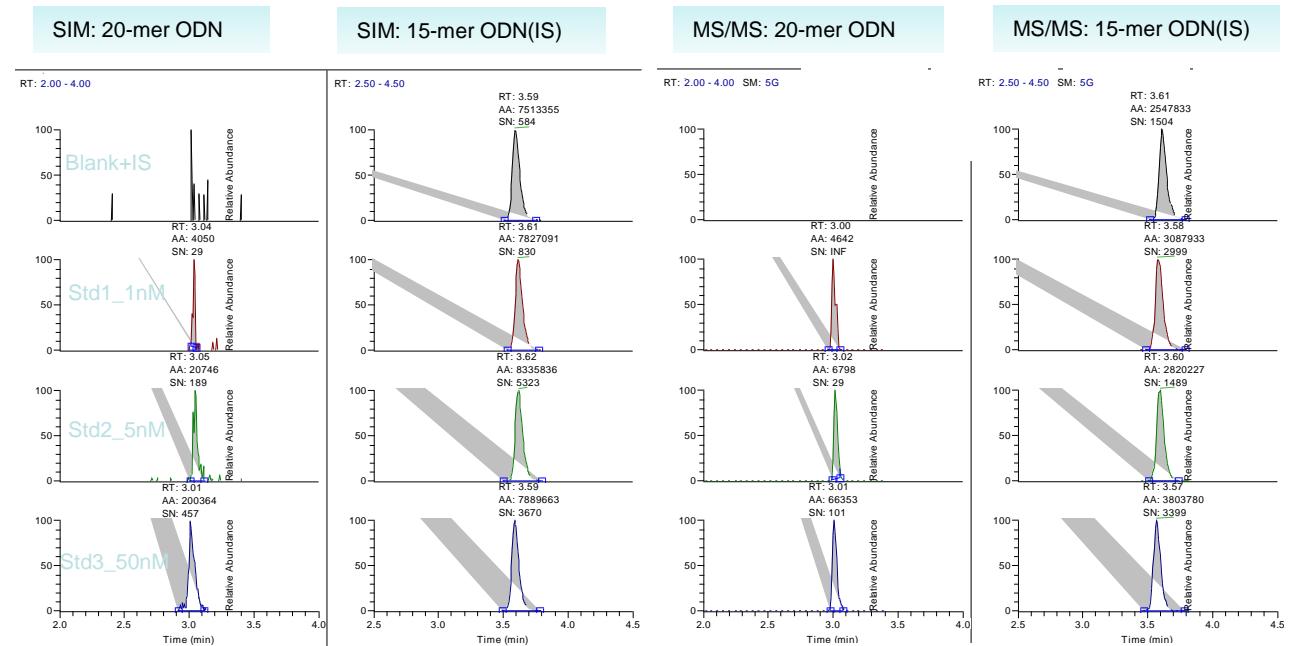


FIGURE 8. Calibration Curve of 20-mer ODN by SIM Method

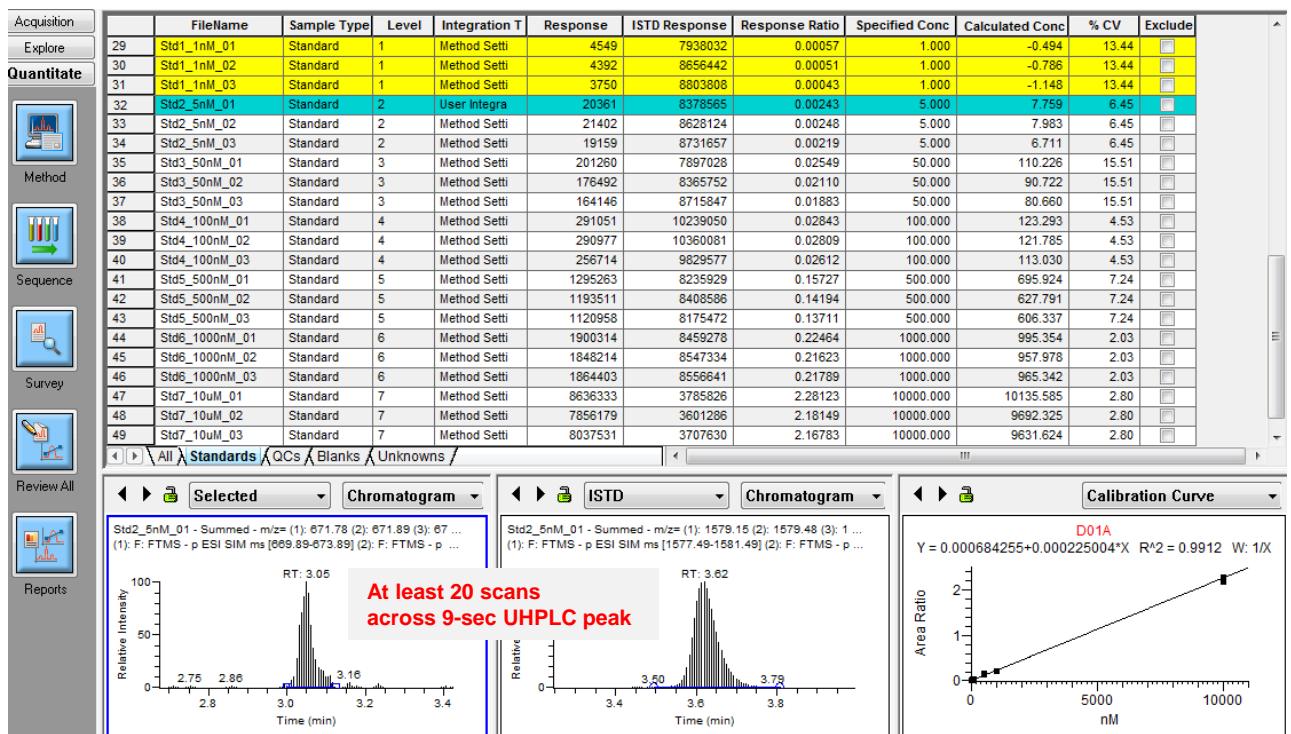
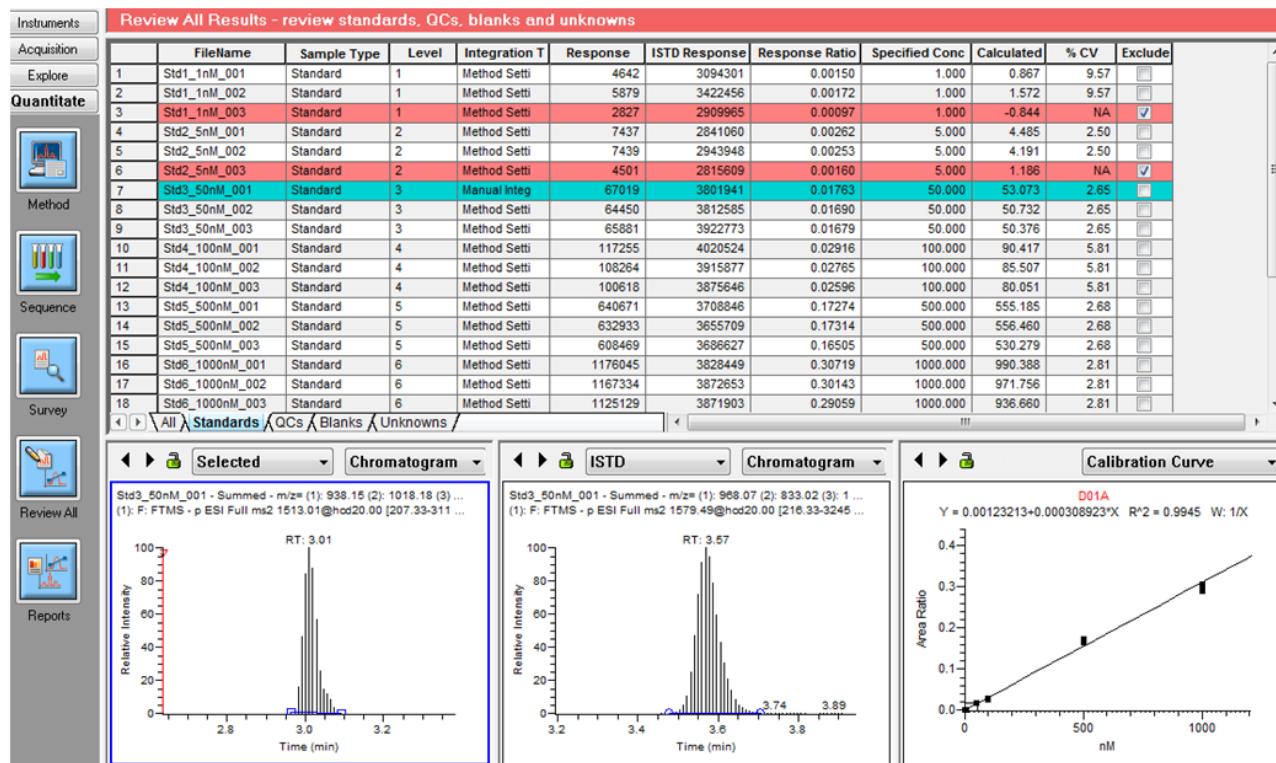


FIGURE 9. Calibration Curve of 20-mer ODN by MS/MS Method



Conclusion

1. Oligonucleotides were identified by high resolution accurate mass data with charge distribution. Their sequences were confirmed by MS/MS accurate mass spectrum. A 20-mer synthetic ODN was used as a model compound and was quantified by both SIM and targeted MS/MS methods.
2. Samples with human plasma as a carrier indicates good linearity with a calibration range of 1-5nM to 10 μ M for SIM method ($R^2=0.9912$, Linear 1/X with CV<16%), while 5nM-1 μ M for MS/MS method ($R^2=0.9945$, Linear 1/X with CV<10%).
3. SIM method (70,000 resolution) shows 5 fold more sensitive than targeted MS/MS method (17,500 resolution) for the synthetic ODN.
4. There are at least 20 scans across a 9-sec UHPLC peak around LOQ at 70,000 resolution.
5. SIM at 140,000 resolution and targeted MS/MS provide better selectivity for the assay with complex matrices.

Reference

1. Wang LX. Oligonucleotide Bioanalysis: Sensitivity versus Specificity. *Bioanalysis*, 2011, 3, 1299-1303.
2. Yuan WW, Wang LX, Enriquez C, Meng M, Wang J, Cook K and Bennett P. Quantitation of Oligonucleotides in Human Plasma Using Q-Exactive Orbitrap High Resolution MS. *Poster presentation at 60th ASMS Conference*, 2012.

©2012 Thermo Fisher Scientific Inc. All rights reserved. ISO is a trademark of the International Standards Organization. This information is presented as an example of the capabilities of Thermo Fisher Scientific Inc. products. It is not intended to encourage use of these products in any manners that might infringe the intellectual property rights of others. Specifications, terms and pricing are subject to change. Not all products are available in all countries. Please consult your local sales representative for details.

Africa-Other +27 11 570 1840
Australia +61 3 9757 4300
Austria +43 1 333 50 34 0
Belgium +32 53 73 42 41
Canada +1 800 530 8447
China +86 10 8419 3588
Denmark +45 70 23 62 60

Europe-Other +43 1 333 50 34 0
Finland/Norway/Sweden
+46 8 556 468 00
France +33 1 60 92 48 00
Germany +49 6103 408 1014
India +91 22 6742 9434
Italy +39 02 950 591

Japan +81 45 453 9100
Latin America +1 561 688 8700
Middle East +43 1 333 50 34 0
Netherlands +31 76 579 55 55
New Zealand +64 9 980 6700
Russia/CIS +43 1 333 50 34 0
South Africa +27 11 570 1840

 ISO 9006
REGISTRED
COMPANY

Thermo Fisher Scientific,
San Jose, CA USA is ISO Certified.

Spain +34 914 845 965
Switzerland +41 61 716 77 00
UK +44 1442 233555
USA +1 800 532 4752

Thermo
SCIENTIFIC
Part of Thermo Fisher Scientific